



Research Article

DOI: 10.36959/771/571

Obtainment of Polyhydroxyalkanoates (PHAs) from Microalgae Supplemented with Agro-Industry Residue Corn Steep Liquor

Páblo Eugênio da Costa e Silva* and Laureen Michelle Houllou



Centro de Tecnologias Estratégicas do Nordeste, Av. Prof. Luís Freire, 1 - Cidade Universitária, Recife - PE, Brazil

Abstract

In this study, it was possible to evaluate the production of polyhydroxyalkanoates (PHAs) from the microalgae *Chlorella vulgaris* and *Tetrademus obliquus*, using 1% and 0.25% (v/v) of agro-industrial Corn Steep Liquor residue, a product from corn processing. The biomass was subjected to mechanical extraction using 4% sodium hypochlorite + chloroform, and the extract was further characterized using Fourier Transform Infrared Spectroscopy (FTIR) analysis. It was possible to observe stretching common to PHAs described in the literature, such as the strong vibration of the carbonyl ester group (C=O) at 1735 cm^{-1} and 1720 cm^{-1} , and the presence of methyl (CH₃) and methylene (CH₂) groups, for *C. Vulgaris* and *T. obliquus*, respectively. Therefore, these indications suggest that both microalgae are potential producers of polyhydroxyalkanoate biopolymers, and have possible applications in the production of bioplastics. This is the first report on the production of polyhydroxyalkanoates from these microalgae, especially the isolated strain, *Tetrademus obliquus*.

Keyword

Bioplastic, Polyester, Polyhydroxyalkanoate, Corn Steep Liquor, *Chlorella vulgaris*, *Tetrademus obliquus*

Introduction

Polyhydroxyalkanoates (PHAs) are biopolymers produced by a range of microorganisms, including microalgae, and will be great replacements for conventional petroleum-derived plastics as they have properties similar to synthetic polymers and are biodegradable [1], capable of forming plastic membranes. Bioplastics obtained from renewable biomass have already been produced from first-generation raw materials such as sugar beet, corn or second-generation raw materials such as lignocellulose materials. Currently, more attention is being paid to production from microorganisms, these-called third generation, which do not compete with human, animal, fresh water or arable land [2,3]. The microbial production of PHAs has been reported for decades, however it has been gaining a lot of attention in recent years, due to the wide variety of applications of these biopolymers in several sectors. However, there is a concern with its production, due to the high costs of organic substrates used for PHA production by heterotrophic bacteria. Thus, the use of photosynthetic microorganisms such as microalgae and cyanobacteria can become quite advantageous, since these microorganisms do not require costly nutritional sources and have minimal nutritional requirements such as in organic sources (CO₂, N, P) and light [4].

Materials and Methods

Microalgae and growing conditions

Two microalgae were used in the experiment and both were cultivated in standard Bold's Basal medium [5]. *Chlorella vulgaris* UTEX 1803 was cultivated at 27 ± 1 °C, constant lighting (3800 lux), constant aeration and supplemented with 1% corn steep liquor (v/v) [6]. *Tetrademus obliquus* was grown at 27 ± 2 °C, constant illumination (3000 lux), constant aeration and supplemented with 0.25% corn steep liquor (v/v) [7]. *C. Vulgaris* is a commercial strain (University of Texas, Austin) and *T.obliquus* was isolated from Açude of Apipucos (Recife, Pernambuco, Brazil, coordinates 8°1'13.08" S; 34°55' 56.51" W).

*Corresponding author: Páblo Eugênio da Costa e Silva, Centro de Tecnologias Estratégicas do Nordeste, Av. Prof. Luís Freire, 1 - Cidade Universitária, Recife - PE, 50740-545, Brazil, Tel: 558-199-505-6544

Accepted: March 09, 2022

Published online: March 11, 2022

Citation: Silva PEC, Houllou LM (2022) Obtainment of Polyhydroxyalkanoates (PHAs) from Microalgae Supplemented with Agro-Industry Residue Corn Steep Liquor. J Bot Res 5(1):138-140

Treatment of corn steep liquor

Corn Steep Liquor was kindly provided by the company Ingredient (Cabo de Santo Agostinho, Pernambuco, Brazil), was treated in accordance with Liggett and Koffler [8], with slight modifications. Initially, Corn Steep Liquor was centrifuged at 8000 rpm for 10 minutes to remove solid particles and then

concentrated NaOH was added to adjust pH 8 and autoclaved at 121 °C for 20 minutes. The agroindustrial residue was then centrifuged again and the supernatant was used on the cultures.

Biopolymer extraction

1.0 g of lyophilized biomass of *C. Vulgaris* (CHLOB) and

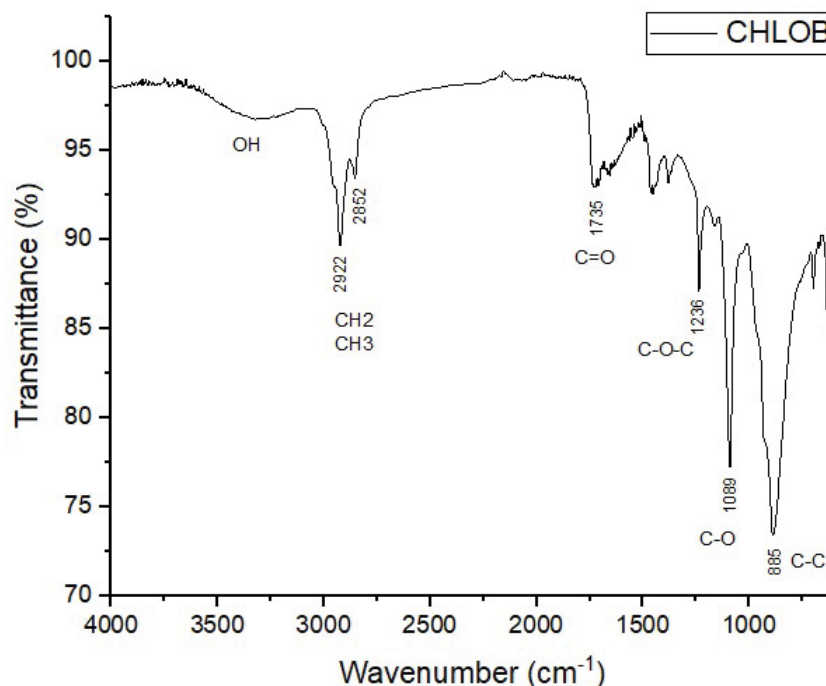


Figure 1: FTIR/ATR spectrum of *C. Vulgaris* extract cultivated in 1% corn steep liquor. The spectrum was recorded ranging from 4000-600 cm⁻¹.

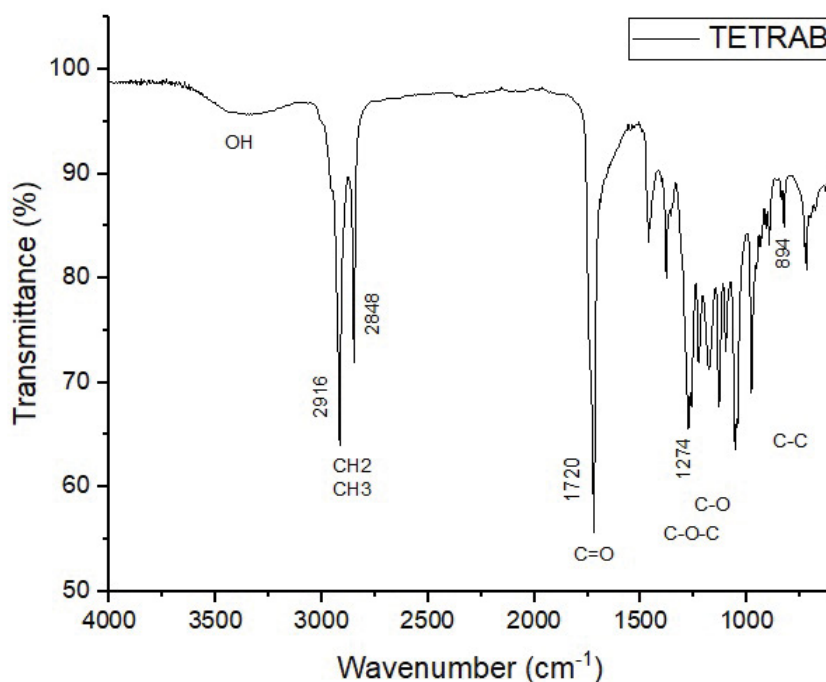


Figure 2: FTIR/ATR spectrum of *T. Obliquus* extract cultivated in 0.25% corn steep liquor. The spectrum was recorded ranging from 4000-600 cm⁻¹.

T. Obliquuos (TETRAB) were subjected to motor agitation with a solution of hypochlorite (4%) + chloroform (v/v), then the samples were centrifuged (3000 rpm, 10 minutes, temperature environment) and the organic phase was collected, evaporated, degreased with hexane and subjected to spectroscopic analysis.

Fourier Transform Infrared Spectroscopy (FTIR)

Samples of PHAs were qualitatively analyzed with Fourier transform infrared spectroscopy (FTIR, Shimadzu, model IR-Tracer 100) between 4000 and 600 cm^{-1} using ATR accessory with a zinc selenide crystal.

Results and Discussion

Fourier Transform Infrared Spectroscopy (FTIR) analysis of PHA samples

Figure 1 and Figure 2 show the spectra of the obtained PHA samples. In the figures it is possible to observe how transmittance bands at 1735 and 1720 cm^{-1} are attributed to strong vibration of the carbonyl ester group (C=O), strongly indicative of medium chain length PHAs (mcl-PHAs), corroborating with Giaquinto, et al. [9] who say that PHBs have bands corresponding to the carbonyl ester group ranging between 1720-1650 cm^{-1} . Other characteristic signs of PHAs are also observed in the spectra, such as the presence of elongations of the -OH group within the carboxyl group, ranging between 3400-3200 cm^{-1} , methyl groups (CH_3) and methylene group (CH_2), with spectra ranging between 2961-2854 cm^{-1} , flash bands between 1466-1000 cm^{-1} are attributed to strong vibration of the C-O group, and other characteristic bands present that are attributed to the C-C group [10,11]. All these indicatives presented as microalgae studied in this work have the ability to synthesize biopolymers capable of forming plastic membranes. Roja, et al. [11] analyzing the elongations obtained in the FTIR of 4 types of algae (*Chlorella* sp., *Oscillatoria salina*, *Leptolyngbya valderiana* and *Synechococcus elongatus*) obtained wave number of the C=O group varying between 1625-1644 cm^{-1} . Costa, et al. [12] evaluating different methods of extraction of the biopolymer obtained from *Spirulina* sp. obtained a strong vibration of the carbonyl ester group (C=O) ranging from 1735-1745 cm^{-1} , corroborating the present study.

Conclusion

The results show that the use of maize for the production of polyhydroxyalkanoates from the microalgae *Chlorella vulgaris* and *Tetrademus obliquus* is possible. In the spectroscopic analysis (FTIR) it was possible to observe the main characteristic elongation of the PHAs, with a strong vibration of the est carbonyl group (C=O), in addition to the methyl (CH_3) and methylene (CH_2) groups. Therefore, microalgae have a strong industrial potential in the production of plastic biofilms.

Acknowledgements

The authors acknowledge supply of Corn Steep Liquor by Ingredion, Cabo de santo Agostinho - Pernambuco, Brazil and the financial support of the Conselho Nacional de Desenvolvimento Científico e Tecnológico - CNPq (Process: 302284/2021-4).

Conflicts of Interest

None.

References

1. Costa SS, Miranda AL, Morais MG, et al. (2019) Microalgae as source of polyhydroxyalkanoates (PHAs) - A review. *International Journal of Biological Macromolecules* 131: 536-547.
2. Ge S, Champagne P (2017) Cultivation of the marine macroalgae chaetomorpha in municipal waste water for nutrient recovery and biomass production. *Environ Sci Technol* 51: 3558-3566.
3. Qiu S, Wang L, Champagne P, et al. (2019) Effects of crystalline nanocellulose on waste water-cultivated microalgal separation and biomass composition. *Applied Energy* 239: 207-217.
4. Afreen R, Tyagi S, Singh GP, et al. (2021) Challenges and perspectives of polyhydroxyalkanoate production from microalgae/cyanobacteria and bacteria as microbial factories: An assessment of hybrid biological system. *Front Bioeng Biotechnol* 9: 624885.
5. Bischoff HW, Bold HC (1963) Phycological studies IV, Some soilalgae from Enchanted Rock and related algal species, University of Texas Publication 6318: 95.
6. Silva PEC, Barros RC, Albuquerque WWC, et al. (2018) In vitro thrombolytic activity of a purified fibrinolytic enzyme from *Chlorellavulgaris*. *Journal of Chromatography B* 1092: 524-529.
7. Souza ATV, Silva PEC, Barros PDS, et al. (2016) Otimização da Produção de Enzimas Fibrinolíticas pela Microalga *Scenedesmus* sp. XII Seminário Brasileiro de Tecnologia Enzimática (ENZITEC).
8. Liggett RW, Koffler H (1948) Corn steep liquor in microbiology. *Bacteriol Rev* 12: 297-311.
9. Giaquinto CDM, Souza GKM, Caetano VC, et al. (2017) Evaluation of the mechanical and thermal properties of PHB/canola oil films. *Polímeros* 27: 201-207.
10. Tanikkul P, Sullivan GL, Sarp S, et al. (2020) Biosynthesis of medium chain length polyhydroxyalkanoates (mcl-PHAs) from palm oil. *Case Studies in Chemical and Environmental Engineering* 2: 100045.
11. Roja K, Ruben Sudhakar D, Anto S, et al. (2019) Extraction and characterization of polyhydroxyalkanoates from marine green alga and cyanobacteria. *Biocatalysis and Agricultural Biotechnology* 22: 01358.
12. Costa SS, Miranda AL, Assis DJ, et al. (2018) Efficacy of *Spirulina* sp. polyhydroxyalkanoates extraction methods and influence on polymer properties and composition. *Algal Research* 33: 231-238.

DOI: 10.36959/771/571